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The Structure of a C₅H₄N₄O₄ Species Trapped by Silylation in Peroxidase Mediated Uricolysis.

A Reactive Ring-Contraction to Spirodihydantoin

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Abstract: Evidence for the species trapped by trimethylsilylation in peroxidase mediated oxidation of urate (1) having the tetrakis(trimethylsilyl)spirodihydantoin (4b) structure is provided by its hydrolysis, reaction with diazomethane, and actual isolation and identification of tetramethylspirodihydantoin (4c).

Following the first report on the stepwise nature of the uricase reaction, a transient UV-absorbing species has been detected sufficiently often in chemical and enzymic oxidations of uric acid (1) to establish it as being of general validity as the initial uricolytic intermediate. This intermediate has hitherto defied all attempts at direct identification owing to its high chemical reactivity. Hence, the assumption, chiefly based on inferential evidence, that 2a is the key uricolytic intermediate has long been awaiting an experimental confirmation. A problematic ring contraction process 2a-3a is often encountered in mechanistic interpretations of the pathway to allantoin. The announcement that 3a can be trapped as 3b by silylation during the initial stage of electrochemical or peroxidase-catalysed oxidation of 15.6 is therefore sufficiently provocative that it can hardly avoid drawing critical attention to the evidence on which it is based. In the absence of a detailed structural information this evidence consists of the mass spectrum of a tetrasilylated species, exhibiting a molecular ion at m/e 472, the most intense peak at m/e 357, and prominent fragments at 457, 342, 329 and 314. In fact, the reported mass spectral data have a suspicious resemblance to those of the tetrasilylated derivative 4b^{7,8} of spirodihydantoin (4a).

To decide whether the silylated species lies on the pathway leading to allantoin or it is merely an alternative end-product of the silylation reaction, the oxidation of 1 with peroxidase [EC 1.11.1.7] was quenched and the UV-absorbing intermediate trapped according to the published procedure,⁶ and the resultant silylation mixture was evaporated *in vacuo*. The residual solid was dissolved in hot water (10 ml), evaporated, and dried (10-4 Torr, 48 h over P_2O_5). This was treated with ethereal diazomethane (2×10 ml); after 48 h ether was removed and the product (R_f 0.16) isolated by preparative TLC (0.5 mm silica gel PF_{254} , CHCl₃). Sublimation (220°C, 10-3 Torr)¹⁰ yielded the pure product (ca 3 mg) identical with the authentic tetramethylspirodihydantoin (4c).¹¹

This simple experiment has been used successfully to demonstrate the stable nature of the basic skeleton of the species trapped by trimethylsilylation. It is obvious that the parent spirodihydantoin (4a) cannot be a uricolytic intermediate, but the spiro-contraction does provide support for 2a as the real intermediate since it is most unlikely that 3a could undergo the sequence which ultimately leads to actual silylated 4b.

In the explanation of the ring-modifying reaction of 2a under silvlation conditions, two general types of mechanism may be discerned. The reaction could clearly involve either [6-4]ring-contraction by shift of carbon or [4-5]ring-contraction by shift of nitrogen, via a hydration equilibrium. No detailed information is available in order to allow a clear distinction between these regiochemical alternatives, which are otherwise reminiscent of ring-contraction processes in flavin pseudobases. Therefore, all the available experimental evidence now seems consistent only with the intermediacy of 2a without recourse to 3a; a coherent uricolytic pathway to allantoin involves ring opening at the 1.6-bond, migration of carboxylate, and decarboxylation as salient steps. 2.3

References and Notes

- 1 For reviews, see Mahler, H. R., "Uricase", in The Enzymes, 2nd ed., Vol. 8, eds. P. D. Boyer et al., Academic Press, N. Y., 1963, pp. 285-296; Vogels, G. D.; Van der Drift, C. Bacteriol. Rev. 1976, 40, 403-468.
- 2 Poje, M.; Sokolić-Maravić, L. Tetrahedron 1986, 42, 747-751; Ibid. 1988, 44, 6723-6728; Sokolić, L.; Modrić, N.; Poje, M. Tetrahedron Lett. 1991, 32, 7477-7480; Modrić, N.; Poje, M.; Watkin, D. J.; Edwards, A. J. Ibid. 1993, 34, 4679-4682.
- 3 Modrić, N.; Derome, A. E.; Ashcroft, S. J. H.; Poje, M. Tetrahedron Lett. 1992, 33, 6691-6694.
- 4 The tetraazabicyclo[3.3.0]octene structure 3a for the incipient intermediate (λ_{max} 305nm) was first proposed by Bentley, R.; Neuberger, A. Biochem. J. 1952, 52, 694-699; Dalgliesh, C. E.; Neuberger, A. J. Chem. Soc. 1954, 3407-3414. Cf. also ref. 1.
- 5 Goyal, R. N.; Brajter-Toth, A.; Dryhurst, G. J. Electroanal. Chem. 1982, 131, 181-202. In earlier papers the trapped species had been regarded as 2b: cf. Brajter-Toth, A.; Dryhurst, G. Ibid. 1981, 122, 205-213. If, instead of quenching the reaction, the initial UV-absorbing intermediate has been allowed to decay away and the resulting solution is lyophilized and then silylated, a pentasilylated derivative of allantoin was detected by its mass spectrum.
- 6 Goyal, R. N.; Brajter-Toth, A.; Dryhurst, G.; Nguyen, N. T. Bioelectrochem. Bioenerg. 1982, 9, 39-60. The procedure adopted by us was following: reaction of 1 (3mM, 4ml) with peroxidase (Sigma, type XII, 80 units/ml, 3ml) in 0.5M NaCl and 5mM phosphate (pH 7.5) was quenched by freezing the mixture in a thin layer (liquid N₂) 100 s after addition of H₂O₂ (10mM, 3ml). This was lyophilized (<0°C, 4 10-4 Torr), and the dry residue heated (125°C, 30 min) with bis(trimethylsilyl)acetamide (3ml) in pyridine (10ml). The procedure was repeated ten times, and the mixtures were combined and used in the next step.
- 7 Tetrakis(trimethylsilyl)spirodihydantoin (4b), m. p. 128-130°C, was prepared in up to 88% isolated yield by heating (130°C, 30 min) powdered 4a (552 mg, 3 mmol) and bis(trimethylsilyl)acetamide (*Merck*, 10ml) in dry pyridine (10ml); after removal of solvent the residue was kugelrohr-distilled (trimethylsilylacetamide: 95°C/10-2 Torr, and 4b: 190°C/10-2 Torr). MS, m/e 474(1), 473(2), 472(4, M+), 459(3), 458(6), 457(14), 359(14), 358(29), 357(100), 344(2), 343(3), 342(8), 331(2), 330(3), 329(8), 316(2), 315(4), 314(9), 301(1), 286(2), 285(5), 270(1), 269(1), 242(2), 241(1), 199(5), 188(3), 174(1), 173(2), 172(2), 171(6), 147(7), 127 (3), 100(10), 73(43). IR (KBr) 1780, 1710 (CO) cm⁻¹. NMR (CDCl₃), 8¹H: 0.45 (s, SiMe₃), 0.32 (s, SiMe₃); 8¹³C 174.5 (s, C-4), 162.3 (s, C-2), 83.6 (s, C-5), -0.9 (q, SiMe₃), -1.1 (q, SiMe₃).
- 8 M. ps, are corrected. IR spectra: Perkin-Elmer FT-IR 1725X; NMR spectra: Varian Gemini-300 (1 H, 300 MHz; 1 C, 75 MHz; in 3 units from internal TMS or CDCl₃). Mass spectra: Varian MAT CH-7, 70 eV, 100 μ A, 3 kV.
- 9 For classical chemistry of spirodihydantoins (4), see Biltz, H. J. Prakt. Chem. 1936, 145, 65-228 (cf. pp. 220-228); it should be emphasized that the bicyclic uric acid glycol structure, originally assigned to the synthetic precursor of 4a, was subsequently revised and shown to be a 3,4-ring-opened isomer: Poje, M.; Paulus, E. F.; Ročić, B. J. Org. Chem. 1980, 45, 65-68.
- 10 The substance was transferred into a tube (6×1 cm) with hot methanol and concentrated at its lowest part by slow evaporation of the solvent. A simple sublimation device, dealing with minute amounts of material, consists of a stout capillary, acting as a receiver, made by drawing out the tube ca. 3 cm above the sample.
- 11 4c, m. p. 228-229°C, was prepared by reaction of 4a with an excess of ethereal diazomethane (cf. ref. 9). MS, m/e 240(40), 225 (2), 211(35), 183 (18), 168(4), 155(66), 154(14), 140(9), 127(2), 126(4), 98(7), 83(14), 70(100). IR (KBr) 1776, 1718 (CO) cm⁻¹. NMR (DMSO- d_6), δ^1 H: 2.99 (s, NMe), 2.77 (s, NMe); δ^1 C: 165.4 (s, C-4), 155.2 (s, C-2), 80.0 (C-5), 25.5 (s, 2NMe).
- 12 Mechanistic precedents are found in formation of their isomeric 10a- and 4a-spirohydantoins: Smith, S. H.; Bruice, T. C. J. Am. Chem. Soc. 1975, 97, 2875-2881; Iwata, M.; Bruice, T. C.; Carrell, H. L.; Glusker, J. P. Ibid. 1980, 102, 5036-5099.